Calyptochloa sphaerocarpa E.J.Thomps. (Poaceae: *Panicoideae*), a new species from central Queensland

E.J. Thompson

Summary

Thompson, E.J. (2021). Calyptochloa sphaerocarpa E.J.Thomps. (Poaceae: Panicoideae), a new species from central Queensland. Austrobaileya 11: 135–154. A new species, Calyptochloa sphaerocarpa E.J.Thomps., endemic to central Queensland is described and illustrated, both for general morphology and for anatomy and surface micromorphology of the leaf and inflorescence culm. It is distinguishable from the other three species of Calyptochloa by characters including the cleistogamous axillary racemes consisting of three to four spherical spikelets with scabrid upper glume and lower lemma, spherical caryopsis and glabrous culm internodes.

Key Words: Poaceae; *Panicoideae*; *Calyptochloa*; *Calyptochloa sphaerocarpa*; Australia flora; Queensland flora; new species; species taxonomy; identification key; anatomy; micromorphology

E.J. Thompson, c/o Queensland Herbarium, Department of Environment and Science, Brisbane Botanic Gardens, Mt Coot-tha Road, Toowong, Queensland 4066, Australia. Email: john.thompson@ des.qld.gov.au

Introduction

Calyptochloa C.E.Hubb. is a genus of three endemic species of Australian grasses, allied to Cleistochloa C.E.Hubb. and Dimorphochloa S.T.Blake and to be included in a new subtribe as circumscribed in Thompson (accepted). The subtribe is defined by having reproductive dimorphism with amphigamous inflorescences and corresponding dimorphic spikelets (Hubbard 1933; Thompson & Simon 2012; Thompson accepted; Thompson & Fabillo 2021; Fig. 1). Calvptochloa is characterised by having a stoloniferous growth habit and dimorphic reproductive system with terminal inflorescences. These consist of spike-like panicles comprising chasmogamous (CH: flowers open to release stigmas and anthers with the potential of cross-pollination) spikelets and axillary racemes with obligate cleistogamous (CL: self-pollination within a closed flower that never opens) spikelets completely or partially hidden by the enveloping leaf sheath. Calyptochloa differs from Cleistochloa by

several characters including the rhizomatous, tufted, growth habit and CL spikelets with an elaiosome (Thompson accepted).

The three species of *Calyptochloa* are consistently grouped with *Cleistochloa* sp. (Duaringa K.B.Addison 42) in various topologies generated from analyses using multiple algorithms and data sets (Thompson accepted; Thompson & Fabillo 2021). This close affinity suggests *Cleistochloa* sp. (Duaringa K.B.Addison 42) should be transferred to *Calyptochloa*.

The purpose of this paper is to formally describe the new species, *Calyptochloa sphaerocarpa* E.J.Thomps., with the phrase name entity *Cleistochloa* sp. (Duaringa K.B.Addison 42) referred to its synonymy.

Materials and methods

This paper is based primarily on plant material held at the Queensland Herbarium (BRI), field collections by the author and material observed in cultivation in Brisbane.

Accepted for publication 19 November 2021, published online 22 December 2021

[©] Queensland Herbarium 2021. ISSN 2653-0139 (Online)



Fig. 1. Glossary of inflorescence components. Del. E.J. Thompson.

Leaf, culm and spikelet materials were obtained from herbarium specimens and fresh material was taken from cultivated plants of the four species of *Calyptochloa*. Nursery stock was initially grown from *ex situ* plants and additional stock was propagated from caryopses and stolons that readily root at the nodes during humid weather in summer. Plants of all species of *Calyptochloa* were cultivated in pots under nursery conditions at latitude 27.5° S, from 2013–2020.

Data used for circumscriptions provided in this paper were obtained from the 161-character list and data matrix provided by Thompson (accepted). Measurements are inclusive, i.e. 1.0-2.7 is given as 1-2.7.

Nomenclature and terminology

Botanical nomenclature follows Thompson (2021a).

General botanical terminology follows Harris & Harris (1994) and Beentje (2010) with additional terminology provided in **Fig. 1**. Terminology relating to inflorescences and spikelets follows Tothill & Hacker (1983), Jacobs *et al.* (2008), Gibson (2009) and Thompson (2021b, accepted). The spikelet equates to a spike in the broader context of inflorescences (Kellogg 2006; Endress 2010). Terminology relating to grass anatomy and micromorphology follows Ellis (1976, 1979), Watson & Dallwitz (1992) and Dengler *et al.* (1994).

Imagery

Photographs were taken using two light microscopes, firstly using a Nikon SMZ25 binocular microscope with Nikon DS-Ri1 camera and images viewed with NIS-Elements BR (ver. BR 5.11.000 64-bit, USA; Laboratory Imaging (http://www.lim.cz, accessed 15 December 2019), and secondly leaves were examined using a Leica DMLB compound binocular microscope with an industrial digital camera and images viewed using ToupView (ver. x64 4.7.14326.20190401, China; Touptek (http://wwwtouptek.com, accessed 20 September 2019).

Thompson, Calyptochloa sphaerocarpa

Scanning electron micrographs (SEMs) were obtained without sputter coating using a Phenom G2 5kev SEM with backscatter detector. Magnifications in **Figs. 6**, **9–14** are those at which the images were taken.

Leaf and inflorescence culm anatomy and surface micromorphology

Transverse sections of leaves and inflorescence culms were obtained using the freehand sectioning method described by Thompson (2017) and modified from Frohlich (1984). Several sections of both leaves and culms were made using fresh material from the cultivated plants.

Leaf surface micromorphology of the abaxial surface was examined using replicas from fresh leaves following the method described by Hilu & Randall (1984).

Observations of leaf and culm anatomy and micromorphology including stomata, silica bodies and microhairs were recorded following descriptions and classifications used by other authors including De Wet (1960), Metcalfe (1960), Twiss *et al.* (1969), Ellis (1979), Renvoize (1987), Watson & Dallwitz (1992), Siqueiros-Delgado & Herrera-Arrieta (1996), Piperno & Pearsall (1998), Krishnan *et al.* (2000), Siqueiros-Delgado (2007), Lu *et al.* (2009) and Jattisha & Sabu (2015).

Spikelet morphology

Observations of micromorphology were made from SEM micrographs of lower and upper lemmas and upper paleas to classify silica bodies, stomata, epidermal long cell walls, microhairs and macrohairs as recorded by Thompson accepted following established descriptions and classifications (Hsu 1965; Jirasek & Jozifova 1968; Ellis 1979; Valdes-Reyna & Hatch 1991; Snow 1996; Acedo & Llamas 2001; Liu *et al.* 2010; Mashau *et al.* 2015; Olonova *et al.* 2016; Neumann *et al.* 2017).

Images of fresh lodicules and stigmas were obtained using light microscopy. Lodicules were classified as plicate or non-plicate (Hsu 1965; Jirasek & Jozifova 1968; Guedes & Dupuy 1976). Stigma macromorphology was classified by the position of emergence from the spikelet, overall shape outline and colour, and micromorphology by characteristics of the lobes including shape of apex, relative length and tilt (Thiele *et al.* 1996).

Caryopsis and embryo morphology including characters relating to hilum, scutellum, epiblast, embryo spermaderm and stylopodium were recorded and classified using established categories (Kennedy 1899; Reeder 1957; Brown 1959, 1960; Watson & Dallwitz 1992; Klak 1994; Kosina 1995; Snow 1998; Liu *et al.* 2005; Liu *et al.* 2015).

Results

The major differences in morphology of the species of *Calyptochloa* are listed in **Table 1**.

Dimorphisms in the CH and CL inflorescences and spikelets vary among the four species, with Calyptochloa johnsoniana E.J.Thomps. & B.K.Simon and C. sphaerocarpa having closer morphological affinities than C. gracillima C.E.Hubb. and C. cylindrosperma. Calyptochloa johnsoniana and C. sphaerocarpa share axillary racemes composed of one or two or as many as four spikelets respectively, with at least the apical one exposed at maturity, while the others are enveloped by the leaf sheath. C. sphaerocarpa also differs by the presence of a third type of inflorescence, reduced spikelike panicles apical on subordinate branches, and comprises CL spikelets that are similar but smaller than the spikelets in the terminal inflorescences (Figs. 2-4).

CH and CL spikelets in *C. sphaerocarpa* also have micromorphological differences in the stigmas and surface pattern on the upper florets. The stigmas differ by size and the shape of the apices of the lobes (**Fig. 5**). Surface pattern on the upper lemmas and paleas differs by the texture of the longitudinal ridges being coarser in the CL upper florets (**Fig. 6**).

The lodicules of *C. sphaerocarpa* have asymmetric shape similar to other taxa in Cleistochloinae and are broadly similar to *C. johnsoniana* (**Fig. 7**).

Austrobaileya 11: 135-154 (2021)



Fig. 2a. Holotype of *Calyptochloa sphaerocarpa* (*Thompson EJT817 & Simon*, BRI). Sheet 1, showing terminal chasmogamous and axillary cleistogamous inflorescences. Photo: Queensland Herbarium.



Fig. 2b. Holotype of *Calyptochloa sphaerocarpa* (*Thompson EJT817 & Simon*, BRI). Sheet 2, showing terminal cleistogamous inflorescences. Photo: Queensland Herbarium.

Key to the species of Calyptochloa

| 1 | Leaf sheath woody and enveloping a single cleistogamous spikelet; fertile |
|----|--|
| | culms disarticulating at nodes |
| 1. | Leaf sheath with a chartaceous margin, racemes more than 1-flowered, |
| | apical spikelet exposed; fertile culms not disarticulating at nodes |
| 2 | Terminal spikelets 3–4.6 mm long; axillary spikelets 3.5–5.5 mm long C. gracillima |

- Terminal spikelets 5 6 mm long; axillary spikelets 6–7.5 mm long. . . C. cylindrosperma
- 3. Axillary racemes 1 or 2-flowered; spikelets lanceolate, glabrous C. johnsoniana

Calyptochloa sphaerocarpa E.J.Thomps., sp. nov.

Similar to *C. johnsoniana* E.J.Thomps. & B.K.Simon but differing by the axillary racemes comprising 3 or 4 scabrid spikelets that are broadly elliptical. **Typus:** Queensland. LEICHHARDT DISTRICT: 17.3 km west of Baralaba, 24 April 2012, *E.J. Thompson EJT817* & *B.K. Simon* (holo: BRI [AQ1017420 comprising 2 sheets]).

Calyptochloa sp. (Duaringa K.P.Addison 42): Thompson & Simon (2012); Simon & Thompson (2013).

Cleistochloa sp. (Duaringa K.P.Addison 42): Thompson (2016); Thompson (2019); Thompson & Fabillo (2021); Thompson (2021a & b).

Illustration: Thompson (2021b: Fig. 1)

Stoloniferous perennial to c. 50 cm tall, rooting at the nodes and with stolons to 6 m long; vegetative culms c. 2 mm wide and up to 4 m long, copiously branched, nodes 6-many. Culm internodes smooth to scabrid. Ligule a fringe of hairs 0.3–0.4 mm long; contraligule c. 0.1 mm long. Leaf sheaths glabrous to hirsute, one margin pilose the other glabrous, margins chartaceous. Leaf blades at mid-culm 3-11 cm long, 3.5-8 mm wide, ultimately disarticulating; glabrous to occasionally hispid, hairs to c. 1.5 mm long; base truncate, apex mucronate; margins undulate on one side, minutely scabrid, thickened, white. Inflorescences of three types with dimorphic spikelets in separate

parts of the plant. Inflorescence type 1 comprising a terminal inflorescence with a reduced panicle, axes 4-9 cm long, 14-30 flowered, on taller culms; branched near base, branches to c. 1.5 cm long, 2-3-flowered. Spikelets overlapping, appressed to rachis, elliptic, adaxial, heteromorphic, largest at apex, 4.8-6.5 mm long (without awn), 1.3-1.6 mm wide; lateral pedicels 0.5-1 mm long, apical pedicels 3-6 mm long. Lower glume lunate, 0.1–0.5 mm long, membranous, apex truncate, hairy. Upper glume elliptic, 4.8-6.5 mm long, margin hyaline, chartaceous, 7-veined, hispid with moderately dense tubercle-based trichomes to 0.5 mm long over body and to 1 mm on margins, apex obtuse. Lower floret sterile; lemma lanceolate, 4-4.8 mm long, margins hyaline, chartaceous, 7-veined, pubescent with scattered tuberclebased trichomes to 1 mm long over body and margins, apex acute to attenuate; palea absent. Upper florets mostly bisexual, some female, chasmogamous; lodicules fanshaped, 0.3-0.5 mm long. Upper lemma body lanceolate, 2.9-4 mm long, margin flat hyaline, cartilaginous, 3-veined, awn 1.2-3 mm long; palea elliptic, 2.9-3.5 mm long, cartilaginous, obscurely 2-veined, apex acute. Anthers 3, equal, chasmogamous 2.5–3.7 mm long. Caryopses $1.6-2.2 \text{ mm} \log \times 1-1.2 \text{ mm}$ wide. Inflorescence type 2 comprising apical racemes on subordinate culms separate from the terminal spike-like panicles, axes 1.8–2.5 cm long, 8-12-flowered (occasionally with 2-flowered branch at base). Spikelets mostly partly overlapping, appressed to rachis, lanceolate, adaxial, heteromorphic, largest at

Taxonomy



Fig. 3. Types of inflorescences of *Calyptochloa sphaerocarpa*. A. terminal spike-like panicle comprising chasmogamous spikelets. B. reduced spike-like panicle comprising cleistogamous spikelets terminal on subordinate culms. C. axillary racemes with cleistogamous spikelets. Scales as shown. All from *Thompson EJT301 et al.* (BRI). Del. E.J. Thompson.



Fig. 4. Three kinds of spikelets of *Calyptochloa sphaerocarpa*. A–G. chasmogamous spikelet from terminal spike-like panicle. A. adaxial view of spikelet showing small lower glume and lower lemma. B. dorsal view of upper glume. C. dorsal view of upper lemma. D. ventral view of upper lemma. E. ventral view of upper palea. F & G. dorsal and ventral views of caryopsis. H–N. cleistogamous spikelet from axillary raceme. H. adaxial view of spikelet showing small lower glume and lower lemma. I. dorsal view of upper glume. J. dorsal view of upper lemma. K. ventral view of upper palea. M–N dorsal and ventral views of caryopsis. O–U. cleistogamous spikelet from apical raceme on subordinate culm. O. adaxial view of spikelet showing small lower glume and lower lemma. P. dorsal view of upper glume. J. dorsal view of upper glume. J. dorsal view of upper glume. J. dorsal view of upper palea. T & U. ventral view of upper glume. S. ventral view of upper glume. P. dorsal view of upper glume. J. dorsal view of upper glume. J. dorsal view of upper glume and lower lemma. B. ventral view of upper glume. J. dorsal view of upper lemma. R. ventral view of upper lemma. S. ventral view of upper palea. T & U. ventral and dorsal view of upper sis. Scales as shown. All from *Thompson EJT301 et al.* (BRI). Del. E.J. Thompson.



Fig. 5. Chasmogamous and cleistogamous stigmas of *Calyptochloa sphaerocarpa*. A. stigma from spikelet from terminal spike-like panicle. B. rehydrated entangled stigmas from cleistogamous spikelet from axillary raceme. C. branch of stigma from terminal spikelet. D. branch of stigma from axillary cleistogamous spikelet. E. caryopsis from cleistogamous spikelet from apical raceme on subordinate culm showing anthers of two sizes (al & a2) and stigmas (s). F. caryopsis from cleistogamous spikelet from axillary raceme showing much reduced stigmas (s) anthers (a). G. terminal spike-like panicle from cultivated plant. Scales as shown. All from *Thompson EJT301 et al.* (BRI). Photos: E.J. Thompson.



Fig. 6. Scanning electron micrographs of surface of upper lemmas of *Calyptochloa sphaerocarpa* (pp papillae; r ridge), $\times 2000$. A. from chasmogamous spikelet. B. from cleistogamous axillary spikelet. All from *Thompson EJT301* et al. (BRI). Photos: E.J. Thompson.

apex, 2.9-3.8 mm long (without awn), 1.2-1.5 mm wide; lateral pedicels 0.5–1.5 mm long, terminal pedicels included on branches 2-5 mm long. Lower glume lunate, 0.1-0.4 mm long, membranous, apex truncate, hairy. Upper glume elliptic, 3.2-4.8 mm long, chartaceous, 7-veined, pubescent with tubercle-based trichomes 0.5 mm long, apex truncate. Lower floret sterile; lemma lanceolate, 2.7-3.6 mm long, chartaceous, 7-veined, pubescent with tubercle-based trichomes to 1 mm long, apex obtuse, revolute; palea absent. Upper florets bisexual, cleistogamous or partially autogamous (anthers enclosed and stigmas exserted); lodicules 2, linear, c. 0.2 mm long. Upper lemma body elliptic, 2.5-3 mm long, cartilaginous, 3-veined, apex revolute, truncate with awn 0.5-1 mm long; palea elliptic, 2.4-3 mm long, cartilaginous, margin hyaline, obscurely 2-veined, apex revolute, acute. Anthers 3, cleistogamous, unequal, 2 at 0.7–0.9 mm long, 1 shorter 1.1– 1.7 mm long; partially autogamous (stigmas exserted), equal, 1.6-2 mm long. Caryopses 1.8-2 mm long, c. 1 mm wide. Inflorescence type 3 comprising axillary racemes, axes 1-3 cm long, 2-4-flowered, 1or 2 exserted from leaf sheath. Basal spikelets sessile, others with pedicels 0.5-4 mm long. Spikelets 4.5-4.8 mm long (without awn), 2-3 mm wide, adaxial. Lower glume lunate, 0.1-0.2 mm long, cartilaginous, apex truncate, minutely pubescent. Upper glume ovate, 3.5–4 mm long, margin hyaline, woody, apex truncate, involute, scabrid with tuberclebased trichomes. Lower floret sterile; lemma lanceolate, 4-4.8 mm long, 1.4-1.6 mm wide, margin hyaline, woody, 7-9-veined, involute, hispid, apex truncate; palea absent. Upper floret fertile; lemma body ovate, 3.5-4 mm long, indurated, papillate, apex convolute, glabrous, margin flat hyaline, 3-veined, awn 0.5–2.6 mm long; palea elliptic, convolute, c. 3 mm long, indurated, glabrous, striate, obscurely 2-veined. Anthers 3, equal, c. 0.3 mm long. Caryopsis pale, 1.6-2.5 mm long, 1.1-1.5 mm wide, ovoid. Figs. 2-7.

Additional specimens examined: Queensland. LEICHHARDT DISTRICT: Gainaford, Duaringa, Apr 1964, *Addison 42* (BRI); 3 miles [5km] E of Duaringa, Apr 1971, *Munroe 15* (BRI); South Blackwater Mine, Laleham, Jan 1986, *Thompson s.n.* (BRI [AQ0399041]);

3 km NW of River View Station, Dec 1998, Ryan 1394 (BRI); Duaringa SF - 15 km NW of Duaringa, Mar 2009, Naske 03/09 (BRI); Blackwater-Rolleston Road, c. 44 km S (by road) from Railway Street, Blackwater, Jul 2011, Menkins ILM 0501 (BRI); Duaringa SF, 8 km W of Duaringa, Dec 2011, Thompson EJT503 (BRI); 16 km W of Baralaba on edge of road, Dec 2011, Thompson EJT510 (BRI); ibid, Dec 2011, Thompson EJT511 (BRI); 35.5 km SSW of Duaringa, Dec 2011, Thompson EJT506 (BRI); Edge of highway, 53 km NW of Clermont, May 2012, Thompson EJT879 & Simon (BRI); Duaringa SF, 12 km W of Duaringa, May 2012, Thompson EJT882 & Simon (BRI); ibid, May 2012, Thompson EJT886 & Simon (BRI); 20.2 km W of Blackwater on edge of Capricorn Highway, Mar 2011, Thompson EJT296, Simon & Edginton (BRI); Edge of Capricorn Highway, 20 km W of Blackwater, May 2014, Thompson EJT1028 (BRI); 8 km E of Bluff on edge of Capricorn Highway in Walton SF, Mar 2011, Thompson EJT301, Simon & Edginton (BRI); 17.3 km W of Blackwater on edge of Capricorn Highway, Mar 2011, Thompson EJT307, Simon & Edginton (BRI); 9.6 km W of Blackwater on edge of Capricorn Highway, Mar 2011, Thompson EJT308, Simon & Edginton (BRI); 16 km W of Bauhinia Downs, Apr 2012, Thompson EJT811 & Simon (BRI; Edge of Gregory Development Road, 53 km NNW of Clermont, May 2012, Thompson EJT883 & Simon (BRI): Duaringa SF, southern side of Capricorn Highway, c. 5 km W of Duaringa, May 2013, Thompson EJT934 & Simon (BRI); Near Wallaroo Siding, c. 12 km W of Duaringa, May 2013, Thompson EJT925 & Simon (BRI); 16 km W of Bauhinia Downs, Apr 2012, Thompson EJT814 & Simon (BRI). PORT CURTIS DISTRICT: Overdeen SF, Jul 2017, Fensham 6684 (BRI). CULTIVATED. Ashgrove, Mar 2017, Thompson MOR817 (BRI).

Distribution and habitat: Calyptochloa sphaerocarpa is endemic to central Oueensland and has been recorded mostly near Duaringa and Blackwater (Map 1). It is frequently a co-dominant in the ground layer under woodland of lancewood (Acacia shirlevi Maiden), and/or bendee (A. catenulata C.T.White), and occasionally in woodland of lemon-scented spotted-gum (Corymbia citriodora var. citriodora (Hook.) K.D.Hill L.A.S.Johnson) within the Regional & Ecosystems 11.7.2 and 11.7.6, respectively (Queensland Government 2020).

Typically, the habitat is shady on lateritic landscapes with undulating to steeply sloping terrain and shallow to skeletal soils (**Fig. 8**). It is commonly sympatric with *Calyptochloa* gracillima var. gracillima, *C. johnsoniana* and *Cleistochloa subjuncea* C.E.Hubb., and other grasses including *Aristida queenslandica* Henrard and *A. caput-medusae* Domin are frequently present.



Fig. 7. Ex situ fresh lodicules showing comparison of A. Calyptochloa sphaerocarpa (Thompson MOR817, BRI). B. C. johnsoniana (Thompson MOR799, BRI). Photos: E.J. Thompson.



Fig. 8. Growth habit of *Calyptochloa sphaerocarpa* and habitat of low woodland of *Acacia shirleyi* (lancewood) on laterite (*Thompson EJT510*, BRI). Photo: E.J. Thompson.



100 µm



Fig. 9. Scanning electron micrographs of surface of lower lemmas of *Calyptochloa sphaerocarpa*. A. from chasmogamous spikelet. B. from cleistogamous axillary spikelet, both \times 500 (bm bicellular microhair; h hook; tbmh macrohair; p prickle; sb silica body; sp spicule-like trichome; tbsh, tuberculate-based simple hair; S stoma). From *Thompson EJT301 et al.* (BRI). Photos: E.J. Thompson.



Fig. 10. Scanning electron micrograph of abaxial leaf surface of *Calyptochloa sphaerocarpa*, $\times 1000$ (acw anticlinal walls; **bm** bicellular microhair; **sb1** silica body – bilobate type; **sb2** silica body – polylobate type; **S** stoma). From *Thompson EJT301 et al.* (BRI). Photo: E.J. Thompson.



Fig. 11. Abaxial leaf surface replica of *Calyptochloa sphaerocarpa*, $\times 20$ (acw anticlinal walls; bm bicellular microhair; h hook; sb1 silica body – bilobate type; sb2 silica body – polylobate type; S stoma). From *Thompson EJT301 et al.* (BRI). Photo: E.J. Thompson.



Fig. 12. Transverse section of fresh leaf at mid vein of *Calyptochloa sphaerocarpa*, ×20. From *Thompson EJT301 et al.* (BRI). Photo: E.J. Thompson.



Fig. 13. Transverse section of portion of fertile culm of *Calyptochloa sphaerocarpa*, ×20 (Vascular bundles: 1 primary; **2** secondary; **3** tertiary). From *Thompson EJT301 et al.* (BRI). Photo: E.J. Thompson.



Fig. 14. Scanning electron micrograph of surface of terminal inflorescence culm of *Calyptochloa sphaerocarpa*, ×500 (**bm** bicellular microhair; **h** hook; **p** prickle; **S** stoma). From *Thompson EJT301 et al.* (BRI). Photo: E.J. Thompson.

Phenology: Flowers December to July.

Conservation Status: Calyptochloa sphaerocarpa has been recorded for four State Forests - Duaringa, Dawson Range, Expedition and Overdeen. Plants are usually common at sites. The species is presently adequately conserved but its response to fire requires research.

Etymology: The species epithet is derived from Latin and refers to the spherical shape of the axillary cleistogamous spikelets.

Breeding system: Calyptochloa has dimorphic breeding system with а amphigamous inflorescences comprising CH terminal spike-like panicles and CL axillary racemes with distinct spikelet dimorphism. Calvptochloa sphaerocarpa differs bv also having occasional apical racemes on subordinate culms comprising spikelets with similar morphology to those in the terminal panicles but smaller. The spikelets in these racemes can be either of two types, completely CL or partially autogamous, with heteromorphic anthers. The partially autogamous spikelets consist of an upper floret with anthers of two sizes enclosed by the lemma and palea and exserted stigmas (**Fig. 5**).

Micromorphology and macromorphology of the lemmas: The CH and CL lower lemmas (**Figs. 6 & 9**) differ in several ways including type of macrohairs, viz. tuberculate-based simple hairs vs tuberculate-based spiculelike trichomes; abundance of stomata, viz. occasional vs absent; and abundance of prickles, viz. common vs abundant. Upper lemmas differ the width of the longitudinal ridges, narrow vs wide.

Abaxial leaf blade epidermis (Figs. 10 & 11): Costal/intercostal zonation conspicuous. Papillae absent. Costal long cells rectangular, much narrower than intercostal; anticlinal walls of intercostal long cells Ω -shaped. Stomata 34–45 µm long with low triangular subsidiaries, in single rows separated by 5–7

Thompson, Calyptochloa sphaerocarpa

files of long cells. Bicellular microhairs $85-90 \mu m$ long, distal cell longer than proximal. Silica bodies predominantly bilobate or occasionally polylobate, $22-28 \mu m$ long. Hooks sparse.

Transverse section of leaf blade (Fig. 12): C,; XyMS+. Mesophyll without radiate chlorenchyma; adaxial palisade inconspicuous. Midrib not prominent; with a double bundle sheath; partial outer ring of clear parenchyma cells with abaxial thick-walled cells and inner ring of partially thick-walled cells with adaxial clear parenchyma cells. Bulliform cells in discrete regular groups, in simple fans. Sclerenchyma accompanying mid-vein as adaxial and abaxial girders; secondary and tertiary vascular bundles as adaxial strands and abaxial girders.

Transverse section of culm (Fig. 13): Culm examined *c*. 1 mm in diameter. Outer smallest vascular bundles adjacent to tangential girder sclerenchyma and imbedded in large-celled sclerenchyma. Vascular bundles with a ring of clear parenchyma; three sizes in separate circles, smallest to the periphery. Chlorenchyma in rectangular blocks, 3–4 cells deep by *c*. 12 cells wide; outer layer of cells more elongated than the more or less circular, inner three layers. Inner ground tissue consisting of large thin-walled cells.

Surface of inflorescence culm (Fig. 14): Scabrid with hooks and prickles. Stomata abundant, 25–25 μ m long, smaller than those on the abaxial leaf surface. Bicellular microhairs, c. 75 μ m long, occasional. Silica bodies absent as for other species of *Calyptochloa* and taxa in *Cleistochloinae*.

Acknowledgements

Much gratitude to Dr G.P. Guymer for provision of resources and equipment at the Queensland Herbarium.

References

ACEDO, C & LLAMAS, F. (2001). Variation of micromorphology characters of lemma and palea in the genus *Bromus* (Poaceae). *Annals Botany Fennici* 38: 1–14.

- BEENTJE, H. (2010). The Kew Plant Glossary an illustrated dictionary of plant terms. Kew Publishing: Kew.
- BROWN, W.V. (1959). The epiblast and coleoptile in the grass embryo. *Bulletin of the Torrey Botanical Club* 86: 13–16.
- (1960). The morphology of the grass embryo. *Phytomorphology* 10: 215–223.
- DENGLER, N.G., DENGLER, R.E., DONNELLY, P.M. & HATTERSLEY, P.W. (1994). Quantitative leaf anatomy of C_3 and C_4 grasses (Poaceae): bundle sheath and mesophyll surface area relationships. *Annals of Botany* 73: 241–255.
- DE WET, J.M. (1960). Culm anatomy in relation to taxonomy. *Bothalia* 7: 311–316.
- ELLIS, R.P. (1976). A procedure for standardizing comparative leaf anatomy in the Poaceae: 1. The leaf-blade as viewed in transverse section. *Bothalia* 12(1): 65–109.
- (1979). A procedure for standardizing comparative leaf anatomy in the Poaceae: 2. The epidermis as seen in surface view. *Bothalia* 12(4): 641– 671.
- ENDRESS, P.K. (2010). Disentangling confusions in inflorescence morphology. *Journal of Systematics and Evolution* 48: 225–239.
- FROHLICH, M.W. (1984). Freehand sectioning with parafilm. *Stain Technology* 59: 61–62.
- GIBSON, D.J. (2009). Grasses & Grassland Ecology. Oxford University Press: Oxford, UK.
- GUEDES, M. & DUPUY, P. (1976). Comparative morphology of lodicules in grasses. *Botanical Journal of the Linnean Society* 73: 317–331.
- HARRIS, J.G. & HARRIS, M.W. (1994). *Plant Identification Terminology: an illustrated glossary*. Spring Lake Publishing: Spring Lake, Utah.
- HILU, K.W. & RANDALL, J.L. (1984). Convenient method for studying grass leaf epidermis. *Taxon* 33: 413–415.
- Hsu, C.C. (1965). The classification of *Panicum* (Gramineae) and its allies with special reference to the characters of lodicule, style-base and lemma. *Journal of the Faculty of Science University of Tokyo, Section III, Botany* 9: 43–150.
- HUBBARD, C.E. (1933). Calyptochloa gracillima C.E.Hubbard. Hooker's Icones Plantarum 33(3210): 1–3.
- JACOBS, S.W.L., WHALLEY, R.D.B. & WHEELER, D.J.B. (2008). *Grasses of New South Wales*. University of New England: Armidale.

- JATTISHA, P.I. & SABU, M. (2015). Foliar phytoliths as an aid to the identification of Paniceae (Panicoideae: Poaceae) grasses in South India. *Webbia* 70: 115–131.
- JIRASEK, V. & JOZIFOVA, M. (1968). Morphology of lodicules, their variability and importance in the taxonomy of the Poaceae family. *Boletin de la Sociedad Argentina de Botánica* 12: 324–349.
- KELLOGG, E.A. (2006). Beyond taxonomy: prospects for understanding morphological diversity in the grasses (Poaceae). *Darwiniana* 44: 7–17.
- KENNEDY, P.B. (1899) The structure of the caryopsis of grasses with reference to their morphology and classification. U.S. Department of Agriculture Division of Agrostology Bulletin 19: 1–14.
- KLAK, C. (1994) Embryo and caryopsis morphology of danthonoid grasses (Arundinoideae: Poaceae): important characters for their systematics? Honours Thesis, University of Cape Town: Rondebosch.
- KOSINA, R. (1995). Remarks on taxonomy of some species of *Elytrigia s.l.* (Triticeae) in the light of embryo morphology. *Acta Societatis Botanicorum Poloniae* 64: 295–302.
- KRISHNAN, S., SAMSON, N.P., RAVICHANDRAN, P., NARASIMHAN, D. & DAYASNANDAN, P. (2000). Phytoliths of Indian grasses and their potential use in identification. *Botanical Journal of the Linnean Society* 132: 241–252.
- LIU, H., HU, X.Y., LIU, Y.X. & LIU, Q. (2015). Caryopsis micromorphology survey of Sorghum (Poaceae)
 – taxonomic implications. South African Journal of Botany 99: 1–11.
- LIU, Q., ZHANG, D.X. & PETERSON, P.M. (2010). Lemma micromorphological characters in the Chloridoideae (Poaceae) optimized on a molecular phylogeny. South African Journal of Botany 76: 196–209.
- LIU, Q., ZHAO, N.-X., HAO, G., HU, X.-Y. & LIU, Y.-X. (2005). Caryopsis morphology of the Chloridoideae (Gramineae) and its systematic implications. *Botanical Journal of the Linnean Society* 148: 57–72.
- LU, H., ZHANG, J., WU, N., LIU, K.-B., XU, D. & LI, Q. (2009). Phytoliths analysis for the discrimination of foxtails millet (*Setaria italica*) and common millet (*Panicum miliaceum*). Plos One 4(2): 1–15.
- MASHAU, A.C., FISH, L. & VAN WYK, A.E. (2015). Taxonomic significance of the abaxial lemma surface in southern African members of *Helictotrichon* (Poaceae). *Bothalia* 45: 1–8.
- METCALFE, C.R. (1960). Anatomy of the Monocotyledons 1. Gramineae. Oxford University Press: London.

- NEUMANN, K., FAHMY, A.G., MULLER-SCHEEBEL, N. & SCHMIDT, M. (2017). Taxonomic, ecological and paleoecological significance of leaf phytoliths in West African grasses. *Quaternary International* 434B: 15–32.
- OLONOVA, M.V., BARKWORTH, M.E. & GUDKOVA, P.D. (2016). Lemma micromorphology and the systematics of Siberian species of *Stipa* (Poaceae). *Nordic Journal of Botany* 34: 322– 334.
- PIPERNO, D.R. & PEARSALL, D.M. (1998). The silica bodies of the tropical American grasses: morphology, taxonomy, and implications for grass systematics and fossil phytolith identification. *Smithsonian Contributions to Botany* 85: 1–40.
- QUEENSLAND GOVERNMENT. (2020). Regional ecosystem descriptions. https://apps.des.qld.gov.au/ regional-ecosystems/, accessed 27 January 2020.
- REEDER, J.R. (1957). The embryo in grass systematics. American Journal of Botany 44: 756–768.
- RENVOIZE, S.A. (1987). A survey of leaf-blade anatomy in grasses XI Paniceae. *Kew Bulletin* 42: 739– 768.
- SIMON, B.K. & THOMPSON, E.J. (2013). Poaceae. In P.D. Bostock & A.E. Holland (eds)., *Census of the Queensland Flora 2012*. https://id.biodiversity. org.au/instance/apni/51389765, accessed 10 November 2021.
- SIQUEIROS-DELGADO, M.E. (2007). Culm anatomy of *Bouteloua* and relatives (Gramineae: Chloridoideae: Boutelouinae). Acta Botanica Mexicana 78: 39–59.
- SIQUEIROS-DELGADO, M.E. & HERRERA-ARRIETA, Y. (1996). Taxonomic value of culm anatomical characters in the species of *Bouteoua* Lagasca (Poaceae: Eragrostoideae). *Phytologia* 81: 124– 141.
- SNOW, N. (1996). The phylogenetic utility of lemmatal micromorphology in *Leptochloa s.l.* and related genera in subtribe Eleusininae (Poaceae, Chloridoideae, Eragrostideae). *Annals of the Missouri Botanical Garden* 83: 504–529.
- (1998). Caryopsis morphology of Leptochloa sensu lato (Poaceae, Chloridoideae). SIDA, Contributions to Botany 18: 271–282.
- THIELE, H.L., CLIFFORD, H.T. & ROGERS, R.W. (1996). Diversity in the grass pistil and its taxonomic significance. *Australian Systematic Botany* 9: 903–912.
- THOMPSON, E.J. (2016). Poaceae. In P.D. Bostock & A.E. Holland (eds.), *Census of the Queensland Flora* 2015. https://id.biodiversity.org.au/instance/ apni/51390821, accessed 10 November 2021.

- (2017). Elionurus purpureus E.J.Thomps. (Panicoideae: Andropogoneae: Rottboelliinae), a new species for Queensland: circumscription and breeding system. Austrobaileya 10: 139– 162.
- (2019). Poaceae. In G.K. Brown & P.D. Bostock (eds.), *Census of the Queensland Flora 2019*. http://data.qld.gov.au/dataset/census-of-thequeensland-flora-2019/, accessed 21 August 2020.
- (2021a). Poaceae. In G.K. Brown & P.D. Bostock (eds) (2021). Census of the Queensland Flora 2020. http://data.qld.gov.au/dataset/census-ofthe-queensland-flora-2020/, accessed 15 May 2021.
- (2021b). A review of the classification and taxonomic and geographic distribution of cleistogamy in Australian grasses. *Australian Journal of Botany* (In press).
- (accepted). Simonachne, a new genus for Australia segregated from Ancistrachne s.1. (Poaceae: Panicoideae: Paniceae) and a new subtribe Cleistochloinae. Australian Systematic Botany.

- THOMPSON, E.J. & SIMON, B.K. (2012). A revision of *Calyptochloa* C.E.Hubb. (Poaceae), with two new species and a new subspecies. *Austrobaileya* 8: 634–652.
- TOTHILL, J.C. & HACKER, J.B. (1983). *The Grasses* of Southern Queensland. University of Queensland Press: St Lucia.
- TWISS, P.C., SUESS, E. & SMITH, R.M. (1969). Morphological classification of grass phytoliths. Soil Science Society of America Journal 31: 109–115.
- VALDES-REYNA, J. & HATCH, S.L. (1991). Lemma micromorphology in the Eragrostoideae (Poaceae). Sida 14: 531–549.
- WATSON, L. & DALLWITZ, M.J. (1992). *The Grass Genera* of the World. University Press: Cambridge.

| Character | Calyptochloa cylindrosperma | Calyptochloa gracillima | Calyptochloa johnsoniana | Calyptochloa sphaerocarpa |
|---|--------------------------------|----------------------------|-----------------------------|------------------------------|
| Leaves: length (cm) \times width (mm) | 1.5-3 ×2-4 | 1.2-4 ×2.5-6 | 2-5.5 ×3-5 | 3-11 ×3.5-8 |
| Culm internodes | pilose | pilose | pilose | glabrous |
| CL racemes apical on subordinate culms | absent | absent | absent | present |
| Number of spikelets in axillary racemes | 1 | 1 | 1 or 2 | 3 or 4 |
| Fertile culm disarticulating | yes | yes | no | no |
| Upper glume and lower lemma of axillary spikelets | smooth | smooth | smooth | scabrid |
| Axillary spikelet length (mm) | 6–7.5 | 3.5-5.5 | 6-6.1 | 4.5-4.8 |
| Outline shape of axillary spikelet | lanceolate | lanceolate | broadly lanceolate | broadly elliptical |

Table 1. Morphological differences between the species of Calyptochloa



Map 1. Distribution of Calyptochloa sphaerocarpa based on Queensland Herbarium collection records.